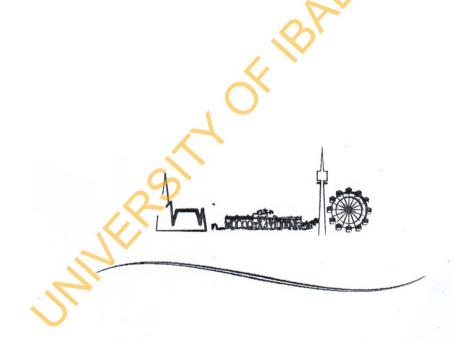
## XV<sup>th</sup> ISAH Congress 2011

July 3 — 7, Vienna, Austria

# Proceedings Volume III



International Society for Animal Hygiene







	2.4.4. A practice oriented three-step basic program against paratuberculosis in cattle Khol, JL; Baumgartner, W	201
	2.4.5. Paratuberculosis control in Austria Geisbauer, E; Altmann, M; Khol, JL; Damoser, J; Österreicher, E; Dünser, M	203
	2.4.6. On the occurrence of Paratuberculosis in cattle and wild animals in Austria/Styria  Hiesel, J; Spergser, J; Deutz, A	205
Sess	sion 2.5. Climate & Air	
	2.5.1. Meat production, climate change and ethics  Gunnarsson, S; Algers, B; Lerner, H; Nordgren, A	209
	2.5.2. Livestock`s "short shadow"? Balancing mitigation of climate change against other values  Lerner, H; Algers, B; Gunnarsson, S; Nordgren, A	213
	2.5.3. Housing emissions of NH <sub>3</sub> , N <sub>2</sub> O and CH <sub>4</sub> and outdoor emissions of CH <sub>4</sub> and N <sub>2</sub> O from organic broilers <i>Meda, B; Hassouna, M; Fléchard, C; Lecomte, M; Germain, K; Picard, S; Cellier, P; Robin, P</i>	215
	2.5.4. Concentrations of airborne particulate matter, ammonia and carbon dioxide in large scale uninsulated lo housing cowsheds in Estonia  Kaasik, A; Maasikmets, M; Aland, A	
	2.5.5. The effect of Rambutan peel (Nephelium lappaceum) as reducing agent on in vitro methane production within creating environment friendly farming  Aditya, S	
Block 3	3 - Tuesday, July 5, 14:00 – 14: 45	
Sess	sion 3.1. Infectious Diseases	
	3.1.1. Seroprevalence of contagious caprine pleuropneumonia in Tigray and Afar, Northern Ethiopia	
	Abera, BH; Eshetu, L; Mengistu, W; Hailesilassie, M	229
	3.1.2. Epidemiological study of canine visceral Leishmaniasis in Syria  Tabbaa, D; El-Ibraheem, J; Turkumani, A	233
	3.1.3. Isolation and prevalence of pathogenic Leptospira interrogans in slaughtered cattle in two abattoirs in Southwestern Nigeria	
	Jagun, AT; Ajayi, OL; Ilugbo, MO; Olugasa, BO	235
Sess	sion 3.2. PLF Poultry	
	3.2.1. MOLDAVI: a model to predict nutrient and energy fluxes from meat poultry production systems Meda, B; Robin, P; Aubert, C; Rigolot, C; Dourmad, JY; Hassouna, M	239
:	3.2.2. Modelling and control of broiler activity  Demmers, TGM; Cao, Y; Parsons, DJ; Gauss, S; Lowe, JC; Wathes, CM	243
	3.2.3. CALORSTA: a tool for design and evaluation of evaporative cooling systems in poultry houses Hassouna, M; Robin, P; Amand, G; de Oliveira, PAV; Aubert, C	247
	3.2.4. Impacts of furnished cage design on cage floor hygiene and egg quality  Huneau-Salaün, A; Guinebretière, M; Huonnic, D; Michel, V	251
Sess	sion 3.3. Horse Health	
	3.3.1. Effect of free exercise in groups on the behaviour of competition horses housed in single stalls Werhahn, H; Hessel, EF; Schulze, H; Van den Weghe, HFA	255
	3.3.2. Expression of the cortisol receptor and ( $11\beta$ -hydroxysteroid dehydrogenase type 1 and 2) in equine testicular and epididymal tissue	
	Herrera-Luna, CV; Budik, S; Aurich, C	259

### ISOLATION AND PREVALENCE OF PATHOGENIC LEPTOSPIRA INTERROGANS IN SLAUGHTERED CATTLE IN TWO ABATTOIRS IN SOUTHWESTERN NIGERIA

Jagun, A.T.<sup>1</sup>, Ajayi, O.L.<sup>2</sup>, Ilugbo, M. O.<sup>2</sup>, Olugasa, B.O.<sup>3</sup>

<sup>1</sup>Department of Veterinary Pathology, University of Ibadan, Nigeria; <sup>2</sup>Department of Veterinary Pathology, University of Agriculture, Abeokuta, Nigeria; <sup>3</sup>Department of Veterinary Public Health and Preventive Medicine, University of Ibadan, Nigeria

#### SUMMARY

Leptospirosis is a waterborne bacterial disease, transmitted to humans through contaminated water, usually by urine of rodents that are chronically infected by the pathogenic strains. The prevalence and incidence of leptospirosis in slaughtered cattle in relation to the risk of exposure of abattoir workers and the public consumers was determined in this study. The objective of this study was to evaluate the prevalence of bovine leptospirosis in slaughtered cattle in southwestern Nigeria. Kidney samples from 108 cattle were examined. Samples were collected from Bodija abattoir in Ibadan, capital city of Oyo State, and Lafenwa abattoir in Abeokuta, capital city of Ogun State, Leptospira was isolated in Ellinghausen-McCullough-Johnson-Harris broth medium Pathogenecity test was carried out in Guinea pigs. Gross and histopathological lesions were observed in their

kidneys. Leptospira species were isolated from 89 (82.4%) out of 108 kidneys from the slaughtered cattle. Twenty (95.2%) out of 21 kidneys and 69 (79.3%) out of 87 kidneys collected from Ibadan and Abeokuta respectively were positive. Only 31 (28.7%) (9 kidneys from Ibadan and 22 from Abeokuta) kidneys showed visible macroscopic changes, while histomorphological changes such as interstitial nephritis, tubular nephrosis and tubular protein cast were observed. Death of guinea pigs that were inoculated occurred within 24 hours to 168 hrs. The isolation of *leptospira interrogans* and the pathology associated with kidneys obtained in this study indicates that cattle slaughter in public abattoir in South-western Nigeria may be sources of exposure and infection to abattoir workers and the public to leptospirosis.

#### INTRODUCTION

Leptospirosis is a zoonosis of ubiquitous distribution, caused by infection with pathogenic *Leptospira* species. The spectrum of disease caused by leptospires is extremely wide, ranging from subclinical infection to a severe syndrome of multiorgan infection with high mortality. The syndrome, icteric leptospirosis with renal failure, was first reported over 100 years ago by Adolf Well in Heidelberg (Levett, 2001)

Documented information on the role of cattle in the epidemiology of leptospirosis in Nigeria is scanty. At

present, there is no specific control strategy against leptospirosis in Nigeria as little is known about the epidemiology of the infection. Cattle, sheep and goats are known in Nigeria to be kept in smallholder units in close proximity with their owners, thus infection with *leptospira* may pose human health hazard. Hence, the present work was designed to determine the prevalence of leptospirosis in cattle, by leptospire isolation in EMJH medium and histopathological changes associated with infected cattle in south-western Nigeria.

#### MATERIALS AND METHODS

This project was carried out in Ibadan and Abeokuta the capital cities of Oyo and Ogun States respectively, in the south western Nigeria. The animals were slaughtered in the central metropolitan abattoirs in Ibadan and Abeokuta where more than 500 and 200 heads of cattle respectively, are slaughtered daily. One-hundred and eight tidney samples from 108 different cattle with unknown epitospirosis history, slaughtered at the abattoirs were selected for the study. Approximately10-15g of kidney sections each was taken for bacteriological and pathological evaluated5.

The Isolation of *Leptospira* was made from direct roculation of two drops of blood in 5 mL of Ellinghausen-McOllough-Johnson-Harris broth medium (EMJH)

(Difco®-USA) with the addition of 10 % of Rabbit serum and 5-fluorouracil (400 mg/L; Sigma®-USA and chloranphenicol (5 mg/L; Sigma®-USA), nalidixic acid (50 mg/L; Inlab®-BR), neomycin (10 mg/L; Sigma®-USA) and vancomycin (10 mg/L; Acros®-USA). Each sample was inoculated into EMJH medium tubes, incubated at room temperature (28-30°C) in the dark and examined under dark field illumination at intervals of 10 days to check for the growth of leptospires for at least three months. The bacteria load was manually counted with a Petroff Hausser counting chamber for experimental infection. Ten guinea pigs of either sex each weighing 150 to 200 grams were inoculated intraperitoneally with 1 ml of randomly selected isolates of the culture leptospiral (1  $\times$  10°). Two normal

guinea pigs were inoculated with EMJH medium as the negative controls.

#### **RESULTS**

Culturally, leptospires were isolated from 89 (82.4%) out of 108 kidneys from the two states. This consist of 20 (95.2%) out of 21 kidneys and 69 (79.3%) out of 87 kidneys collected from Ibadan and Abeokuta respectively.

The specific histopathology lesions observed in samples that tested either positive or negative for leptospires in EMJH expressed in percentages are shown in table 1.

Out of the one hundred and eight (n = 108) kidney samples randomly collected from the two abattoirs, only 31 (28.7%) (9 kidneys from Ibadan and 22 from Abeokuta) kidneys showed visible macroscopic changes. The lesions include multifocal necrotic areas, multifocal petechiat ions, icterus and diffuse nephrosis

Most of the guinea pigs that were inoculated with the isolates died between 24 and 168hrs (seven guinea pigs) and the remaining three showed signs of infection associated with leptospirosis.

Table 1: Prevalence of specific histopathological lesion in kidney tissue samples collected from Abeokuta and Ibadan that were cultured in EMJH nedium.

lium. Histological lesions		Abeokuta abattoir (n=87)		Ibadan abattoir (n=21)	
		Positive (n=69)	Negative (n=18)	Positive (n=20)	Negative (n=1)
Interstitial Oedema		8 (11.6%)	-	1 (5%)	-
Tubular Nephrosis	i .	52 (75.4%)	2 (11.1%)	16(80%)	1 (100%)
Tubular Epithelail\`.suo	blation	47 (68.1%)	4 (22.2)	6 (30%)	-
Interstitial Fibrosis	1	31 (44.9%)	1 (5.5%)	8 (40%)	-
Interstitial Mononuclea	r Cells Infiltration	58 (84%)	1 (5.5%)	13(65%)	1 (100%)
Perivascular Mononucle	ear Cell Infiltration	38 (55.1%)		7 (35%)	1 (100%)
Periglomerular Mononu	uclear Cell Infiltration	32 (46.4%)	-	3 (15%)	-
Glomerular Atrophy	S	25 (36.2%)	-	4 (20%)	
Cast	<del>\</del>	40 (58.0%)	13(72.2%)	11(55%)	1 (100%)
Tubular Dilation		30 (43.5%)	5 (27.8%)	4 (20%)	1 (100%)
Glomerulonephritis	Membranous	10 (14.5%)	-	1 (5%)	-
	Membranoproliferative	19 (25.5%)	-	6 (30%)	-
Crystals		31 (44.9%)	4 (22.2%)	6 (30%)	1 (100%)

#### DISCUSSION

Leptospires were isolated from 89 out of 108 kidney samples collected from the two abattoirs in the Southwestern Nigeria. The diagnosis was based on either

or both isolation of the leptospira species with EMJH medium at 27°C-30°C and pathogenicity test with guinea pigs. In Nigeria, there is no record of the isolation of

leptospires from animals, as the majority of the data are based on serology(Agunloye et al 1997).

The gross renal lesions, such as cortical haemorrhage, multifocal necrosis, diffuse palor and icterus, reported in this study are typical of renal bovine leptospirosis and are consistent with those in previous reported cases in cattle and other animals (Faine *et al.*, 1999). In this study, there is no relationship between gross lesions and isolation of leptospira organism in the kidney samples, since most of the kidney samples without gross lesions were culturally

positive. Histological changes observed in this investigation were in correlation with the reports of other workers (Marinho *et al* 2009). However, Skilbeck *et al* (1988) did not observe significant histopathological lesions in kidneys from which leptospires were isolated. In this study the lesions range from locally extensive cellular infiltrates to diffuse lesions, characterized by tubular nephrosis, glomerular atrophy and renal haemorrhage. Most of the kidneys samples studied presented changes suggestive of leptospirosis in the histopathological lesions of the kidneys in accordance with Faine *et al.*, (1999).

#### CONCLUSION

The isolation of *leptospira interrogans* and the pathology associated with the kidneys obtained in this study indicate that cattle slaughtered in the public abattoir in the Southwestern Nigeria may be sources of the infectious agent to

human population. It is recommended to improve on the sanitation and personal hygiene of abattoir workers and implement a hazard analysis critical control system in the abattoir.

#### REFERENCES

- AGUNLOYE, C. A., OYEYEMI, M. O.; AKUSU, M. O.; AJALA, O.O. and AGBEDE, S. A.(1997): Clinical and serological diagnosis of leptospirosis in aborting West African Dwarf goats. Bull. Anim. Health. Prod. Afr. 45: 5-8.
- 2. FAINE, S., ADLER, B.; BOLIN, C.; PEROLAT, P. (1999): Leptospira and leptospirosis, 2nd ed. MedSci, Melbourne, Australia.

JANVER SILA

- LEVETT, P. N., BRANCH, S. L.; WHITTINGTON, C.U.; EDWARDS, C.N.; PAXTON. (2001) Two methods for rapid serological diagnosis of acute leptospirosis. Clin. Diagn. Lab. Immunol. 8:349-351.
- MARINHO, M., OLIVEIRA-JUNIOR, I. S; MONTEIRO, C. M. R; PERRI, S. H; SALOMAO, R. (2009): Pulmonary Disease in Hamsters Infected with Leptospira interrogans: Histopathologic Findings and Cytokine mRNA Expressions. *Am J Trop Med Hya* 80: 832-836
- 5. SKILBECK, N. W., and G. T. MILLER. (1986): A serological survey of leptospirosis in Gippsland dairy farmers. Med. J. Aust. 144:565-567.