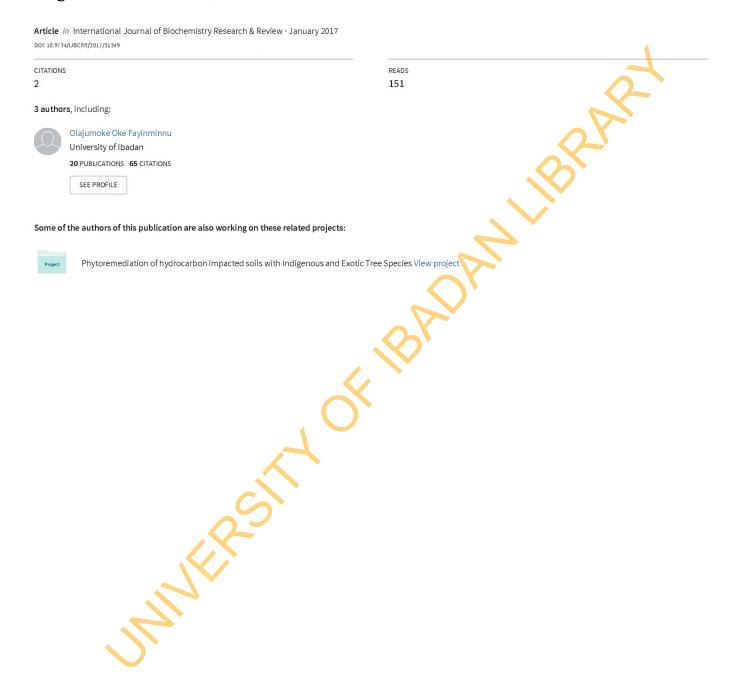
Toxicity Assessment of Sub Lethal Doses of Chlorpyrifos on the Kidney and Liver Organs of Male Wistar Rats





International Journal of Biochemistry Research & Review

17(3): 1-15, 2017; Article no.IJBCRR.31349

ISSN: 2231-086X, NLM ID: 101654445

Toxicity Assessment of Sub Lethal Doses of Chlorpyrifos on the Kidney and Liver Organs of Male Wistar Rats

O. O. Fayinminnu^{1*}, S. O. Tijani¹ and O. O. Fadina¹

¹Department of Crop Protection and Environmental Biology, University of <mark>Ibadan, I</mark>badan, Nigeria.

Authors' contributions

This work was carried out in collaboration between all authors. Author OOF designed the study, wrote the protocol and the first draft of the manuscript. Authors OOF and SOT performed the laboratory trials as well as the statistical analysis. Author OOF reviewed the experimental design and results. Author O. O. Fadina managed the literature searches and edited the manuscript. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJBCRR/2017/31349

Editor(s):

(1) Chunying Li, Department of Biochemistry and Molecular Biology Wayne State University School of Medicine, Detroit, USA.

Reviewers:

(1) Salisu Maiwada Abubakar, Bayero University, Kano, Nigeria.

(2) Hassan I. El-Sayyad, Mansoura University, Egypt.

Complete Peer review History: http://www.sciencedomain.org/review-history/19572

Original Research Article

Received 31st December 2016 Accepted 5th February 2017 Published 16th June 2017

ABSTRACT

Chlorpyrifos, an organophosphate pesticide is an important neurotoxic and tissue damage agent. It is one of the most heavily used pesticides in domestic and agricultural applications globally. Repeated doses of chlorpyrifos have been able to cause significant disturbances on the biochemical and physiological functions of the blood, and histological abnormalities in livers and kidneys exposed to this insecticide. The toxicities of sub lethal oral administration of chlorpyrifos daily for 28 days were assessed using a completely randomized design. Twenty five albino Wistar rats weighing between 150-200 g divided into five groups containing five rats each were housed in the Central Animal house of College of Medicine, University of Ibadan. Chlorpyrifos at 0 (control), 18.9, 25.9, 32.2 and 39.2 mg/kg were orally administered to male rats, respectively for four weeks, between the months of May and June, 2014. At the end of the experimental period, the toxicities of chlorpyrifos were assessed in rats using haematology, serum liver enzymes and histopathological assays. Results revealed significant reduction in body weights compared to control. The packed cell volume (PCV), hemoglobin (Hb) and lymphocytes (Lymp) also showed significant reduction at

39.2, 25.9 and 32.2 mg/kg (39.33, 45.40 and 44.80%), respectively compared to control (49.67%). Insecticide administration to rats resulted in significant elevation of serum transaminases (AST and ALT) and alkaline phosphatase (ALP) from 32.2, 39.2 and 25.9 mg/kg dosages, (ranged from 294.94-542.00 u/L for AST and from 96.25-130.77 u/L for ALT), respectively. Also, experimental treated groups exhibited marked of total protein and altered albumin and globulin contents compared to control. Studies revealed dose dependent increase of histopathological alterations. The livers showed moderate vacuolar change of hepatocytes, having a finely reticulated cytoplasmic and congestion of central veins. The kidneys showed mild focal sloughing off of tubular epithelium of renal cortex, fluid in tubular lumen, tubules appearing dilated and cystic (nephrosis) and proteinaeceous fluid in Bowman's capsule and compressing the glomerulus tufts (Esinophilic). Different concentrations of chlorpyrifos including the lowest tested dose produced marked alterations in the exposed animals in this study and thereby affecting the overall performance in terms of health and wellbeing. Thus, this could cause similar health and environmental risks to humans even at the lowest dose.

Keywords: Chlorpyrifos; toxicity; biochemical; haematology; histopathology.

1. INTRODUCTION

Chemical pesticides are to improve the quantity and quality of food to feed the ever increasing world human and animal population and promote public health [1]. However, they become one of most widely available environmental contaminants that are deliberately released to the environment. Pesticides fall into numerous chemical classes, which have widely differing biological activities and thus differing potential to produce toxic effects in living organisms, including humans [2] and their poisonings in developing countries had been occurring as a result of their misuse. Unintended exposure to toxic pesticides can occur during manufacturing, formulation and application or from environmental residues after application [3]. Chemical plant workers, transport workers and pesticide applicators (especially the farmers) [4] are the major occupational groups that might be exposed to different toxic pesticides. Also workers in industrial settings are at increased risk since they handle various toxic chemicals including pesticides, raw materials, toxic solvents and inert carriers.

Organophosphates (OP's) are the esters of pentavalent phosphorous acid, exhibiting wide range of toxicity in mammals [5]. Organophosphate insecticides are one of the most widely used chemicals in agriculture and public health [6]. The compelling needs to improve human and animal nutrition and promote public health have led to increase in organophosphates usage in recent time as they are used extensively to control agricultural, household and structural pests [7].

Chlorpyrifos, (O, O-diethyl-O-(3,5,6-trichloro-2pyridyl}-phosphorothioate) is a chlorinated organophosphate (OP) insecticide with a broad spectrum of activities such as acaricide and nematicide used on pests of plants, animals and humans. It has been widely used in domestic, agricultural and to control public health pests such as mosquitoes and fire ants [8]. Chlorpyrifos has been reported as one of the most widely used organophosphate insecticides, thereby accounting for more than 50% of global insecticidal use [9]. Chlorpyrifos' toxicity is typically manifested in the central and peripheral svstem where acetylcholinesterase by its active metabolite (Chlorpyrifos- oxon) leading to cholinergic hyper stimulation. Another associated mechanism implicated in the induced toxicosis includes the induction of oxidative stress the bν organophosphate compounds [10].

Literatures have reported about sub chronic toxicological studies of chlorpyrifos in rats which revealed reduced weight gain and slight histopathological changes in adrenal gland with significant brain and plasma AChE inhibition [11]. The evidence that chlorpyrifos causes alteration histopathological, changes in some hematological and biochemical parameters in experimental animals were reported from the studies of Kazmi et al. [12-14]. Chlorpyrifos, an organophosphate has also been identified in showing a number of additional toxic effects which include hepatic dysfunction, haematological and immunological abnormalities, embryotoxicity, genotoxicity and neurobehavioral changes as reported by the earlier researchers: Heikal et al. [15-19].

Humans and animals are occasionally and unintentionally exposed to lethal and sub-lethal doses of pesticides [20]. This stems from its continuous uses in-door, due to household use in the control of termites and as protectants in the preservation of kola, maize and yam flour chips, pesticide exterminator application and dietary (ingestion) exposure [21] thereby aggravating toxicity in human and ill health. Previous researchers have reported that repeated exposure to chlorpyrifos has been shown to cause severe damage to the vital organs. Therefore, due to the widely usage of chlorpyrifos domestically, in agriculture and public health continuously necessitated the study of its toxicological effects. This present study was aimed at assessing the toxicity of sub lethal doses of Chlorpyrifos on haematology, biochemical and histopathology of kidney and liver organs of male wistar rats.

2. MATERIALS and METHODS

2.1 Experimental Sites

The experiment was carried out in the Department of Crop Protection Environmental Biology (CPEB) Toxicology laboratory, Faculty of Agriculture and Forestry; Central Animal house of College of Medicine, Clinical pathology Histopathology and Laboratories of the Department of Veterinary Pathology, Faculty of Veterinary Medicine, all in the University of Ibadan, Nigeria.

2.2 Experimental Animals

Twenty five male Rattus norvegicus (wistar) rats of five weeks old weighing 150 - 200 g used for this experiment; were obtained from the experimental animal house of the Department of Veterinary Anatomy, Faculty of Veterinary Medicine, University of Ibadan, Animals were kept under and maintained at 27±2°C, with 12 hour light, 12 hour dark cycles, relative humidity of 75-80% at the Central Animal house of College of Medicine, University of Ibadan, They were kept in procured, cleaned individual plastic cages for four weeks (28 days). The rats were divided into five groups (A, B, C, D and E) of five rats each in a cage, and were acclimatised for one week before experimentation. Growers mash feed sourced locally and water were given to the animals during acclimatisation and treatment periods ad libitum. All animals were handled in accordance with the guidelines for care and use of laboratory animals as stated by

the Animal care and use research ethic committee.

2.3 Chlorpyrifos Pesticide

Chlorpyrifos trade names include DursbanTM and LorsbanTM [22], 480 g/I EC (Dursban) used for this study was purchased from Saro Agroscience chemicals located at Oluyole Estate Ibadan, Oyo State, Nigeria. All other chemicals required for biochemical, haematological and histopathological estimations were of high quality and were provided by the Department of Pathology, Veterinary Medicine, University of Ibadan.

2.4 Chlorpyrifos Pesticide Sub Lethal Dilution

The Chlorpyrifos 480 g/l stock solution used for this study is liquid and soluble in water. Dilution was carried out on the stock solution using the chemical formula as described by [23] at the Department of Crop Protection and Environmental Biology Toxicology Research Laboratory.

 $C_1V_1 = C_2V_2$

Where C₁ = Initial Concentration of Stock

 V_1 = Volume of Stock used

C₂ = Concentration desired

 V_2 = Volume desired

Dilution of 100 g/l was made from the stock solution.

The next dilution was made from 100 g/l to prepare 10 g/l of the insecticide. The following concentrations of 18.9 mg/l, 25.9 mg/l, 32.2 mg/l and 39.2 mg/l were prepared from 10g/l.

The treatments were 0 mg/kg (Control), 18.9 mg/kg, 25.9 mg/kg, 32.2 mg/kg and 39.2 mg/kg of chlorpyrifos. All the treatments were administered daily. Oral administration of the treatment doses was done through gavaging for 28 days (four weeks).

2.5 Experimental Design/ Animal Grouping

The experimental used was a completely randomized design. The 25 male wistar rats were grouped into five groups of five rats per cage. Groups A B, C, D and E for chlorpyrifos were administered at 0 mg/kg, 18.9 mg/kg, 25.9 mg/kg, 32.2 mg/kg and 39.2 mg/kg, respectively.

2.6 Experimental Toxicological Study

A total of 25 male albino (wistar rats) were used and divided into five groups with five rats in each cage. The various levels of Chlorpyrifos formed the treatments with a control group (0 mg/kg; No pesticide). The treatment levels were 18.9 mg/kg, 25.9 mg/kg, 32.2 mg/kg and 39.2 mg/kg. All the treatments were administered daily (early in the morning) before feeding by oral gavage using a 2 ml oral cannula for a period of 28 days. Weights of the animals were taken weekly and at the end of the experiment.

Observations on toxicity were made on the behavioural patterns of the rats during dosing. These include shyness, signs of pesticide poisoning such as loss of appetite, tremor, jerking of the muscles, constricted pupils, aggressiveness, skin irritation, convulsion, vomiting, weight gain / loss, paralysis and death [24].

On the termination day, 2 ml of blood samples was collected through the ocular orbit from all the surviving rats into vaccum tainer EDTAK3 tubes by heparinized capillary tubes, according to [25]. The blood samples for serum biochemical analysis were collected by heparinized capillary tubes into lithium heparinized tubes. The rats were sacrificed through cervical dislocation and organs of interest were harvested which include the kidney and liver for histopathological examination.

The haematological and serum biochemistry analyses and histopathological examinations were carried out by the technology experts in the Clinical pathology and Histopathology laboratories of the Department of Veterinary Pathology, Faculty of Veterinary Medicine, University of Ibadan, Nigeria.

2.7 Haematological and Serum Biochemical Analyses

Blood samples collected for packed cell volume (PCV) and Haemoglobin (Hb) count were determined using the micro haematocrit and cyanmethemoglobin methods, respectively as described by [25]. The Red blood cells (RBCs) and White blood cells (WBCs) were determined using improved Neubauer haemocytometer after appropriate dilution [26]. The differential lymphocyte counts were determined by scanning Giemsa's stained slides in the classic manner [26]. Blood chemistry analysis was also done to ascertain Total protein (TP) using the serum total

as estimated by Biuret method [27]. Alkaline Phosphatase (ALP), Alanine aminotransferase (ALT) and Aspartate aminotransferase (AST) serum were obtained using the procedure of [28].

2.8 Histopathological Study

Preparation of experimental animal tissues (kidney and liver organs) of each rat for microscopic examination, the histopathological procedures were followed in a stepwise protocol such as fixation, dehydration, clearing, infiltration. embedding, blocking, sectioning and staining. Tissue specimens were collected from rats' livers and kidneys and rapidly fixed in 10% neutral buffered formalin. After proper fixation, thin paraffin sections were routinely prepared and stained with Hematoxylin and Eosin (H & E) stain for microscopical examination according to [29] through the light microscope (400x). The histopathological examination was conducted in Department of Veterinary Pathology, Faculty of Veterinary Medicine, University of Ibadan, Nigeria.

2.9 Statistical Analysis

Data obtained in this study were analysed with one way analysis of variance (ANOVA), group means were compared for significance at 95% confidence level (p< 0.05) by Fischer's Least Significant Difference (LSD). Results were presented as mean ± standard error of the mean (SEM).

3. RESULTS

3.1 Behavioural/Morphological Responses

Paralysis as a morphological response of chlorpyrifos toxic effects was observed among the experimental animals dosed with 32.2 mg/kg treatment at week three in this present study.

3.2 Feed Intake and Weight Gain/Loss

Effects of different concentrations of chlorpyrifos on the feed intake of wistar rats from this study showed significant differences (p< 0.05) as presented in Fig.1. Treated rats with 25.9 mg/kg dose of chlorpyrifos had the least feed intake (72.0%) followed by treatment 18.9 mg/kg in having (73.0%). Feed intake of experimental rats dosed with 39.2 mg/kg chlorpyrifos also decreased progressively (82.0%). However, animals from the control (0 mg/kg) had significant

higher feed intake (96.0%) over other treatments. All the animals treated reduced progressively on feed intake with no significant difference (p> 0.05) from each other except from the control. The results on toxicity of different concentrations of chlorpyrifos on body weights showed the treated animals with no significant difference (p> 0.05) amongst each other from week zero (0) to week three (3). However, a highly significant difference (p<0.05) between the control (212.0 g) and treated animals was observed (Fig. 2).

3.3 Heamatological Observations

The results showed the toxic effects of chlorpyrifos on hematological parameters: packed cell volume (PCV) and lymphocytes showed significant differences (p< 0.05), while the neutrophiles and monocytes revealed

no significant difference (p>0.05) as shown in Fig. 3.

The PVC revealed no significant difference (p> 0.05) between the treatments 18.9 mg/kg, 25.9 mg/kg, 32.2 mg/kg and control, while there was a significant difference (p< 0.05) between treated animals of chlorpyrifos at 39.2 mg/kg and the control. Experimental rats from the control had significant higher mean value (49.67%) for PVC. while the lowest mean value (39.33%) was obtained from animals treated with 39.2 mg/kg chlorpyrifos (Fig. 3). The lymphocytes showed no significant difference (p> 0.05) between the control and treated animals with 18.9 mg/kg, 25.9 mg/kg and 39.2 mg/kg. However, 32.2 mg/kg treatment differed significantly (p<0.05) when compared with the control. The control had the highest mean value (62.33%)the lymphocytes, while animals treated

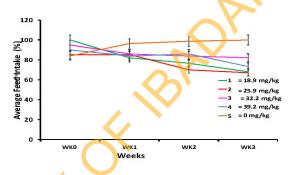


Fig. 1. Effects of varying treatments of chlorpyrifos on feed intake of wistar rats

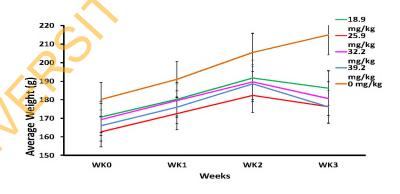


Fig. 2. Effects of varying treatments of Chlorpyrifos on the average weekly weight (g) gain of wistar rats

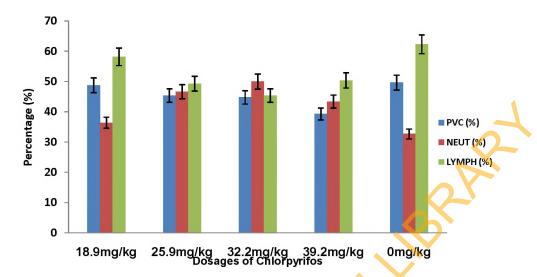


Fig. 3. Effects of varying levels of sub lethal of Chlorpyrifos on haematological parameters of wistar rats

PVC = Packed Cell Volume, NEUT= Neutrophiles, LYMPH = Lymphocytes

32.2 mg/kg chlorpyrifos had the lowest value (45.40%) (Fig. 3). No significant differences (p>0.05) were observed amongst all the treated and control animals in the neutrophiles and monocytes of haematological parameters. However, the neutrophiles had highest mean value (50.00%) in animals treated with 32.2 mg/kg and lowest value (32.67%) in the control. The trend in monocytes revealed highest mean value (6.20%) in animals treated with 39.2 mg/kg and lowest (4.00%) in 25.9 mg/kg (Fig. 3).

The hemoglobin results in this study showed no significant difference (p> 0.05) from animals treated with 18.9 mg/kg, 32.2mg/kg, 39.2 mg/kg (15.6, 14.6 and 14.8 g/dl) chlorpyrifos and control (15.9 g/dl). While, the animals treated with 25.9 mg/kg differed significantly (p< 0.05) in having (12.6 g/dl) from 18.9 mg/kg (15.6 g/dl) and the controlled animals (15.9 g/dl). The highest mean

(15.9 g/dl) for haemoglobin was recorded from the control while lowest value (12.6 g/dl) was from 25.9 mg/kg chlorpyrifos treated animals (Fig. 4).

The results on toxic effects of chlorpyrifos on WBCs revealed no significant difference (p> 0.05) amongst all the treatments and control. Although the control had highest mean value (8300x10⁶/mm³) WBCs, for while experimental animals treated with 18.9 mg/kg chlorpyrifos had the lowest mean value (4460 x10⁶/mm³) (Fig. 5). No significant difference (p>0.05) was observed amongst all the treated and control animals in the RBCs haematological parameter. However, animals treated with 32.2 mg/kg had highest mean value (5.80x10⁶/mm³) and lowest (3.10x10⁶/mm³) was from 25.9 mg/kg in RBCs as shown in Fig. 6.

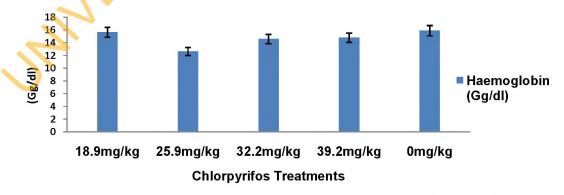


Fig. 4. Effects of varying levels of sub lethal of Chlorpyrifos on Hemoglobin level of wistar rats

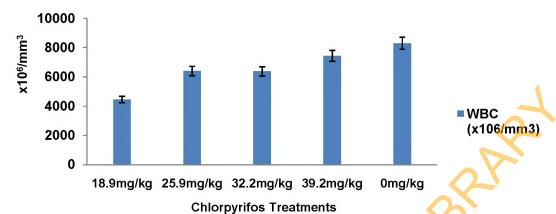


Fig. 5. Effects of varying levels of sub lethal of Chlorpyrifos on White Blood Cell Counts of wistar rats

WBC = White Blood Cell Counts

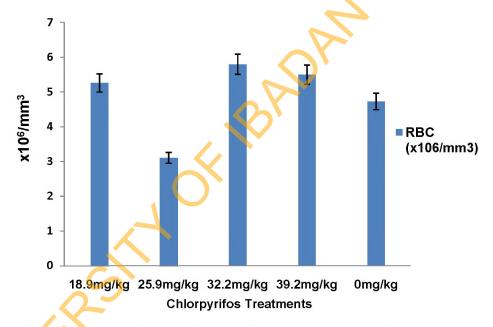


Fig. 6. Effects of varying levels of sub lethal of Chlorpyrifos on Red Blood Cell Counts of wistar rats

RBC= Red Blood Cell counts

3.4 Serum Biochemistry Observations

Results of toxic effects of chlorpyrifos on serum biochemical parameters on experimental animals revealed significant differences (p<0.05) in this study.

There was a significant difference (p< 0.05) in alkaline phosphatase (ALP) of rats treated with 25.9 mg/kg chlorpyrifos and control. Although no different significance (p>0.05) was observed amongst the other treatments and the control.

However, the treatment 25.9 mg/kg had the highest mean value (417.9 u/L), while the lowest (173.97 u/L) was from 32.2 mg/kg treatment (Fig. 7.). The aspartate amino transferase (AST) results on experimental rats showed that, there were significant differences (p<0.05) between the treatments 18.9 mg/kg and 25.9 mg/kg when compared with the control. No significant differences (p>0.05) were observed between the control and the chlorpyrifos treatments 32.2 mg/kg and 39.2 mg/kg. Although, 32.2 mg/kg treated animals had highest mean value

(542.23u/L), while 18.9 mg/kg chlorpyrifos had the lowest (262.00 u/L) (Fig. 7). Significant difference (p< 0.05) was observed in Alanine amino transferase (ALT) between 39.2 mg/kg treatment and control animals. Significant differences (p<0.05) were also shown between 39.2 mg/kg and 18.9 mg/kg chlorpyrifos treated animals as well as 25.9 mg/kg treatment. Meanwhile, no significance differences (p>0.05)were observed between 32.2 25.9 mg/kg, mg/kg and 18.9 mg/kg chlorpyrifos treated animals when compared with control Although, the animals. 39.2 mg/kg treatment had the highest mean value (130.77 u/L), while control 0 mg/kg had the lowest mean value (83.70 u/L) as shown in Fig. 7.

There were no significant differences (p> 0.05) in the total and direct bilirubins in all the chlorpyrifos treatments when compared with the control as presented in Fig. 8. However, in total bilirubin 39.2 mg/kg treatment had the highest mean value (0.57 mg/dL) and the same lowest value (0.47 mg/dL) was from the control and other treatments. In direct bilirubin, 32.2 mg/kg treatment had highest mean value (0.20 mg/dL) while the lowest mean value (0.1 mg/dL) was from 18.9 mg/kg.

The results from toxic effects of different concentrations of chlorpyrifos on total protein, albumin and globulin on the experimental animals showed significant differences (p< 0.05) as presented in Fig. 9.

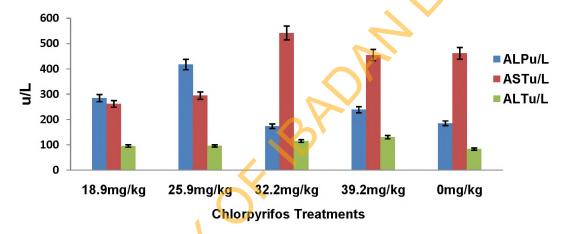


Fig. 7. Effects of varying levels of sub lethal of Chlorpyrifos on serum biochemical parameters of wistar rats

ALP = Alkaline Phosphate, AST=Aspertate Amino Transferase, ALT=Alanine Amino Transferase

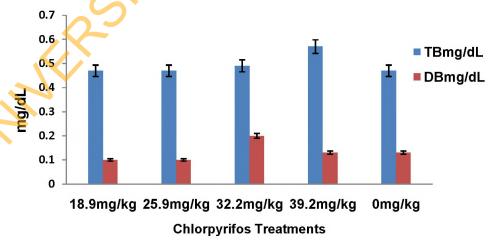


Fig. 8. Effects of varying levels of sub lethal of chlorpyrifos on Total Bilirubin and Direct Bilirubin levels of wistar rats

TB=Total Bilirubin, DB = Direct Bilirubin

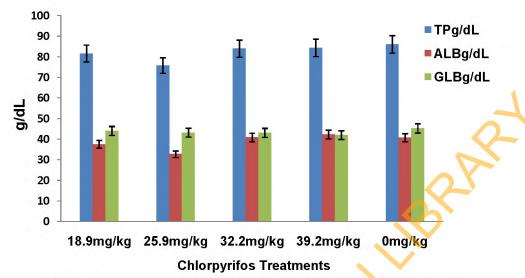


Fig. 9. Effects of varying levels of sub lethal of Chlorpyrifos on serum biochemical parameters of wistar rats

TP=Total Protein, ALB = Albumin, GLB= Globulin

The trend in total protein estimation showed significant differences (p<0.05) between 18.9 mg/kg and 25.9 mg/kg chlorpyrifos treated animals and the control. However, 32.2 mg/kg and 39.2 mg/kg chlorpyrifos treatments showed no significant difference (p>0.05) with the control. The highest mean value (86.10 g/dL) was obtained from the control, while the lowest mean value (75.78 g/dL) was from 25.9 mg/kg chlorpyrifos treated animals (Fig. 9). The result for albumin in the experimental animals showed significant difference (p< 0.05) between 25.9 mg/kg chlorpyrifos and the control, while other treatments 18.9 mg/kg, 32.2 mg/kg and 39.2 mg/kg showed no significant difference (p> 0.05) with the control. The highest mean value (42.30 g/dL) was observed in 39.2 mg/kg chlorpyrifos treated animals and the lowest mean value (32.79 g/dL) was from 25.9 mg/kg treatment (Fig. 9). Globulin ratio result revealed no significant differences (p> 0.05) amongst all chlorpyrifos treated animals and the control. However, highest mean value (45.31 g/dL) was obtained from the control and lowest mean value (42.06 g/dL) was from 39.2 mg/kg treatment (Fig. 9).

Results on the toxic effects of chlorpyrifos on the histopathological examinations (Photomicrographs sections) carried out on the experimental rats (*Rattus norvegicus*) were shown in Plates 1- 2.

The 0 mg/kg dosage (control) in organs examined (liver and kidney) showed no visible

lesions with closely packed hepatic plates in the liver. There were no apparent morphological changes in the livers and kidneys of the control rats. The control group showed no symptoms of any gross abnormalities (Plates 1 and 2).

Observations from 18.9 mg/kg chlorpyrifos treated experimental rats showed that, the animals had mild multifocal sloughing off of tubular epithelium in renal cortex and cortico medullary junction of the kidney. The kidney of experimental rats exposed to 25.9 mg/kg dosage chlorpyrifos expressed widespread presence of eosinophilic (proteinaeceous) fluid in tubular lumens; multifocal moderate sloughing off of tubular epithelial cells, while 32.2 mg/kg dosage of chlorpyrifos showed nephrosis. The kidney of the animals exposed to 39.2 mg/kg dosage of chlorpyrifos showed eosinophilic (proteinaeceous fluid in the Bowman's capsule and compressing the glomeruli tufts) (Plate 1).

The animals treated with 18.9 mg/kg and 25.9 mg/kg chlorpyrifos showed moderate to severe multifocal thinning of hepatic plates in the liver. The histopathological examination result from 32.2 mg/kg chlorpyrifos treated animals showed that, the hepatocytes in the livers were finely reticulated with cytoplasmic appearance. However, there were moderate widespread of vacuolar changes of hepatocytes and congestion of central veins in the livers of some rats exposed to 39.2 mg/kg dosage of chlorpyrifos (Plate 2).

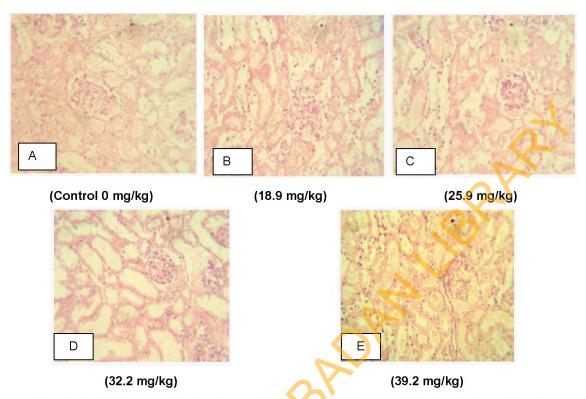


Plate 1. Photomicrographs sections of kidneys of *Rattus norvegicus* groups exposed to different sub lethal concentrations of chlorpyrifos and control

A= No treatment of chlorpyrifos (control 0 mg/kg). No visible lesions. B = Treated with 18.9 mg/kg of chlorpyrifos. Mild multifocal sloughing off of tubular epithelium in renal cortex and corticomedullary junction. C = Treated with 25.9 mg/kg of chlorpyrifos. Widespread presence of eosinophilic (proteinaceous) fluid in tubular lumen; multifocal moderate sloughing off of tubular epithelial cells. D = Treated with 32.2 mg/kg of chlorpyrifos. Multiple foci of moderate sloughing off of tubular epithelial cells; some tubules appearing dilated and cystic (nephrosis). E = Treated with 39.2 mg/kg of chlorpyrifos. Esinophilic (proteinaeceous fluid in Bowman's capsule and compressing the glomeruli tufts). (Hematoxylin and Eosin (H &E), 400x)

4. DISCUSSION

The widespread and extensive use of organophosphate pesticides such as chlorpyrifos (CPF) in developing nations raises the likelihood of inadvertent exposure to the pesticide either from short-term or long-term exposures with consequent toxic effects [30]. In most of these countries, the magnitude of pesticide residue in food of both plant and animal origin is largely unknown, hence, increasing the risk of exposure [31] and ill health.

In this study, paralysis observed on the treated animals could be due to the chlorpyrifos toxicity during dosing which resulted in abnormal morphology [24]. Toxic effects of chlorpyrifos caused a decrease in body weights of rats exposed to different concentrations of the pesticide. Although, the effects did not show in the first week of administration to the animals, but it became apparent on the body weights from

the second week to the third week of treatment with respect to dosages administered at 18.9 mg/kg to 39.2 mg/kg. All experimental rats treated with chlorpyrifos showed a marked decrease in body weights, while untreated (control) rats continued to gain weight progressively during the period of treatments.

This result agreed with [32] who concluded that chlorpyrifos caused a significant change in rats' body weights. The obtained result also conformed with [33] findings that, there were adverse effects on exposure to chlorpyrifos on male rats with a significant decrease in body weights. Jaiswal and Verma [34-36] also reported that the most profound rat stress was the reduction of body weight after administration of chlorpyrifos. Studies conducted by [37-39] gave decreased body weights in rats treated with chlorpyrifos with no effects on physical or reflex development.

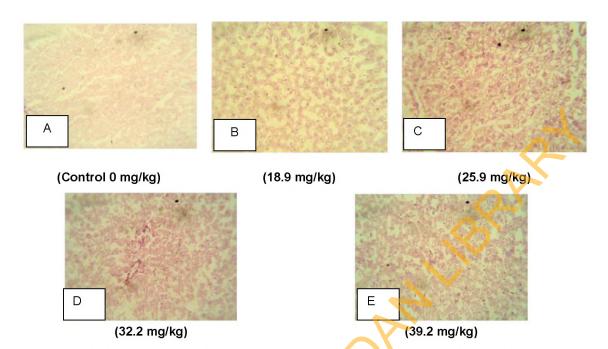


Plate 2. Photomicrographs sections of livers of *Rattus norvegicus* groups exposed to different sub lethal concentrations of chlorpyrifos and control

A= Treated with 0 mg/kg of chlorpyrifos (control). Closely packed hepatic plates with no visible lesions. B = Treated with18.9 mg/kg of chlorpyrifos. Moderate vacuolar change of hepatocytes; severe multifocal thinning of hepatic plates. C = Treated with 25.9 mg/kg of chlorpyrifos. Widespread moderate vacuolar change of hepatocytes; moderate thinning of hepatic cords. D = Treated with 32.2 mg/kg of chlorpyrifos. Multifocal mild thinning of hepatic plates; hepatocytes have a finely reticulated cytoplasmic appearance. E = Treated with 39.2 mg/kg of chlorpyrifos. Moderate widespread vauolar change of hepatocytes and congestion of central veins. (H &E, 400x)

This present study also revealed leucopenia in packed cell volume (PCV) which was apparently lymphopenia, neutropenia, monocytopenia in the chlorpyrifos animal treated groups. The marked reduction in packed cell volume and heamoglobin in the rats' population exposed to varying concentration of chlorpyrifos dosages 39.2 mg/kg, 32.2 mg/kg, 25.9 mg/kg and 18.9 mg/kg resulted in leucopenia and low hemoglobin counts. These findings corroborated previous studies of [6-40] that attributed chlorpyrifos induced leucopenia to neutropenia and lymphopenia. Ambali et al. [6] also reported neutrophilia following chlorpyrifos exposure, in contrary to neutropenia recorded present study. [41] attributed monocytopenia recorded in workers exposed to chlorpyrifos experienced inhibition of monocyte esterase, (alpha)-naphthyl butyrate esterase.

It was shown in this present study that chlorpyrifos caused a decrease in the white blood cell in the rats' population exposed to it with marked reductions observed in 39.2 mg/kg

and 32.2 mg/kg treatments, respectively. This induced immune toxicity is in support of [42] and [43] that chlorpyrifos has been shown to induce immune toxicity in rats either via the induction of apoptosis or necrosis. Chlorpyrifos exposure has been shown to induce immune toxicity via the induction of apoptosis partly mediated through the activation of caspase as reported by [44]. This was found in this study as differential leucocytes which were greatly reduced thereby making the rats susceptible to varying degree of abnormalities like paralysis observed in treatment 32.2 mg/kg. Chronic chlorpyrifos exposure has been associated with abnormality of the immune system including depression of Tlymphocytes [45].

Immunotoxicity of white blood cell (WBC) in chlorpyrifos has been associated with either inhibition of serine hydrolases or esterases in components of the immune system. through oxidative damage to immune organs, by modulation of signal transduction pathways controlling functions [46].

The reduction of serum levels of protein for all the rats dosed with the chlorpyrifos at all dosages of treatment correlates with the findings of [47], this indicated that toxicants may cause stress-mediated mobilization of protein to cope with the detrimental condition so imposed. The protein mobilized is one of the strategies employed to meet the energy required to sustain increased physical activity, biotransformation and excretion of the toxicants. Aminotransferases (ALT and AST) are produced in the liver and are good markers of damage to liver cells but not necessarily the severity of the damage [48,49]. The reduced levels of AST, ALB and ALT in chlorpyrifos treated groups at 39.2 mg/kg, 32.2 mg/kg and 25.9 mg/kg could be as a result of suppression of production by the liver. They are normally present at low levels in the blood so if the liver cells are damaged, it would be expected that some of the enzymes leak into the blood and increase the levels [14]. Also, apparently earlier damaging effect before progression to cell death and leakage of enzymes.

In contrast to serum enzymes, albumin apart from being a useful indicator of the integrity of glomerular membrane is also an important indicator in determining the severity of disease [50,51]. Decrease albumin may be due primarily to reduction in synthesis by the liver and secondarily to reduced protein intake which further confirms hepatic damage [52,53] as observed with 39.2 mg/kg and 32.2 mg/kg in the study. Decreased albumin observed in this work revealed that chlorpyrifos administration may be attributed to hepatic dysfunction.

The chlorpyrifos administered orally to the animals treated showed degeneration in some of renal corpuscles, sloughing off of tubular epithelium, congestion of glomeruli and compression in size of glomeruli tufts of the kidneys. These descriptions agree with the studies of those described by [54-56,14]. The toxicity of chlorpyrifos on the kidneys indicated a dose dependency.

There was loss of cell architecture and increased degeneration of hepatic cells in the livers of the experimental animals administered with chlorpyrifos. Most of hepatic cells were necrotic and the cytoplasm was vacuolar with denoting degenerating of hepatocytes. This finding collaborates with the studies of [57,56]. The changes in treated animal livers indicated that the degeneration of livers increased with the doses of chlorpyrifos to exposure which could be

related to dose-dependent increase in the liver function enzymes.

The toxic effects of different sub doses of chlorpyrifos on livers and kidneys on treated animals in this study is also in agreement with the report from the work of [58] that administration of sub lethal doses of chlorpyrifos resulted in altered enzyme activities of liver and renal (kidney) damage to experimental animals.

5. CONCLUSION

In this study, it has shown that, the chlorpyrifos pesticide at 18.9 mg/kg, 25.9 mg/kg, 32.2 mg/kg and 39.2 mg/kg had toxic effects on all the treated experimental animals which showed decrease in their body weights. Chlorpyrifos at sub lethal doses showed lower values in haematological and biochemical parameters which resulted into stress and paralysis in this study. The reduced values of the serum enzymes (ALT, AST and ALB) at 25.9 mg/kg, 32.2 mg/kg and 39.2 mg/kg chlorpyrifos resulted into liver damages through the leaking of these enzymes into the blood and thereby increasing the levels. The lower level of albumin at 39.2mg/kg chlorpyrifos also contributed to the hepatic dysfunction. The histopathology results in this study revealed serious damaged kidney and liver organs with hepatocytes and nephrosis on animals treated with 32.2 mg/kg and 39.2 mg/kg chlorpyrifos, repectively. However, chlorpyrifos treatments at 18.9 mg/kg and 25.9 mg/kg showed mild to moderate damages in this study.

The different sub lethal concentrations of chlorpyrifos in the treated animal groups in this present study relative to the control group affected their overall performances in terms of health and well being. From the outcome of this study, the haematological and biochemical parameters and histopathology could be used as bio indicators of exposure and effects in instances where chlorpyrifos is used as a pesticide.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

 Gianessi L. The value of insecticides in U.S. crop production. Crop Life Foundation; 2009.

- Available: http://www.croplifefoundation.org/lnsecticide Benefits/Executive Summary (Accessed April 21, 2009)
- Timothy CM, Ballantyne B Pesticide Toxicology and International Regulation. England: John Wiley and Sons Ltd; 2004.
- Jeyaratnam J. Acute pesticide poisoning: A major global health problem. World Health Stat Q. 1990;43(3):139-44.
- 4. Forget G. Pesticides and the third world. Journal of Toxicology and Environmental Health, Part A Current Issues. 1991; 32(1):11-31.
- Sultatos LG. Mammalian toxicology of organophosphorous pesticides. J Toxicol. Environ. Hlth. 1994;43:271-289.
- Ambali, SF, Onukak C, Idris SB, Yaqub LS, Shitu M, Aliyu H, Kawu MU. Vitamin C attenuates short-term hematological and biochemical alterations induced by acute chlorpyrifos exposure in Wistar rats. J. Medici. Med. Sci. 2010;1(10):465-477.
- 7. Pope C, Chaudhuri J. Chakraborti T. Upregulation of muscarinic receptors in rat brain by the organophosphate (OP) insecticide chlorpyrifos (CPF). 23rd annual meeting of the society for neuroscience, Washington DC, USA, Society for Neuroscience Abstracts. 1993;19(13):429.
- 8. US. Environmental Protection Agency (USEPA). Interim reregistration eligibility decision (IRED) Document for dichlorvos (DDVP). Office of Pesticide Programs. Washington, DC. 2006;167.
- Casida JE, Quistad GB. Organophosphate toxicology: Safety aspects of non acetylcholinesterase secondary targets. Chem. Res. Toxicol. 2004;17:983-998.
 Qiao D, Seidler FJ, Slotkin TA. Oxidative
- Qiao D, Seidler FJ, Slotkin TA. Oxidative mechanisms contributing to developmental neurotoxicity of Nicotine and Chlopirifos. Toxicol, Appl. Pharmacol. 2005;206:17-26.
- Zayed SMAD, Faraghaly M, El-Maghraby S. Fate of 14C- chlorpyrifos in stored soybeans and its toxicological potential to mice. Fd. Chem. Toxicol. 2002;41:767-772.
- Kazmi F, Shakoori AR, Hafeez MA, Ali SS. Short-term effects of chlorpyrifos on hematology and biochemical components of blood of the Sprague-Dawley rats. Pak. J. Zool. 2003;35:245-257.
- Jacobson H, Ostergaard G, Lam HR, Poulsen ME, Frandsen H, Ladefoged O, Meyer O. Repeated dose 28-day oral toxicity study in Wistar rats with a mixture of five pesticides often found as residues in

- food: Alphacypermethrin, bromopropylate, carbendazim, chlorpyrifos and mancozeb. Food and Chemical Toxicology. 2004; 42(8):1269-1277.
- 14. Sharma A, Sodhi S. Apminder S. Brar AS, Jyotika K. Ghai JK. Chlorpyrifos-induced hematological, biochemical and histoarchitectural alterations in albino mice. International Journal of Advanced Research. 2013;1(10):825-835.
- Heikal TM, Mossa AH. Marei GIK, Abdel Rasoul MA. Cyromazine and chlorpyrifos induced renal toxicity in rats: The Ameliorated Effects of Green Tea Extract. J Environ Ana Toxicol; 2012a. Available: http://dx.doi.org/10.4172/2161-0525.1000146
- 16. Heikal TM, El-Sherbiny M. Hassan SA, Arafa A, Ghanem HZ. Antioxidant effect of selenium on hepatotoxicity induced by chlorpyrifos in male rats. Int J Pharm Pharm Sci. 2012b;4(4):603-609.
- 17. Mehta A, Verma RS, Srivastava N. Chlorpyrifos induced alterations in the levels of hydrogen peroxide, nitrate and nitrite in rat brain and liver. Pest Biochem Physio.I 2009;94:55–59.
- 18. Qiao D, Seidler FJ. Slotkin TA. Oxidative mechanisms contributing to the developmental neurotoxicity of nicotine and chlorpyrifos. Toxicol Appl Pharmacol. 2005;206:17–26.
- 19. Verma RS, Mehta A, Srivastava N. *In vivo* chlorpyrifos oxidative stress: Attenuation by antioxidant vitamins. Pest Biochem Physiol. 2007;88:191–196.
- 20. Martin B, Stephaine K. William W. Kim MB. Kathy P. Lorrie B. Carol R. Childhood pesticide exposures in the Texas-Mexico border: Clinical manifestations and poison centre use. Am. J. Public Health. 2003; 93(8):1310-1315.
- 21. Dey KR. Choudhury P, Dutta BK. Impact of pesticide use on the health of farmers: A study in Barak valley, Assam (India). J. Environ. Chem. and Ecotoxicol. 2013; 5(10):269-277.
- 22. Beyond Pesticides; 2000. Available: www.beyondpesticides.org
- 23. Oshoke FI, Fadina OO, Fayinminnu OO. Sub-chronic toxicity study of synthetic Pyrethroids (Lambda-cyhalothrin) on reproductive organs of male Wistar rats. Int. J. Adv. Agric. Res. 2016;4:38-45.
- 24. Fadina OO, Taiwo VO, Ogunsanmi AO. Single and repetitive oral administration of

- common pesticides and alcohol on rabbits. Trop. Vet. 1999;17:97-106.
- 25. Dacie SJ, Lewis SM. Basic hematological techniques in practical hematology. International student 6th ed. Churchill Livingstone. Edinburgh, London, Melbourne and New York; 1986.
- Schalm OW, Jain NC, Carroll EJ. Veterinary hematology. 3rd Lee Aand Febriger, Philadelphia. Indian Vet. J. 1975; 39:157-161.
- Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal. Biochem. 1976; 72:248-254.
- Reitman S. Frankel SA. Colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. Am. J. Clin. Pathol. 1957; 28:56–63.
- 29. Drury R, Wallington E. Carlton's histological technique 4th Ed., Oxford University. Press, NY, Toronto; 1986.
- Tang J, Cao Y, Rose LR, Brimfield AA, Dai JA, Goldstein, Hodgson E. Metabolism of chlorpyrifos by human cytochrome P450 isoforms and human, mouse, and rat liver microsomes. Dru. Metab. Dispos. 2001; 29(9):1201-1204.
- 31. Ambali SF. Ameliorative effects of vitamins C and E on neurotoxicological, haematological and biochemical changes induced by chronic chlorpyrifos in Wistar rats. PhD Dissertation, Ahmadu Bello University, Zaria, Nigeria, 2009.
- 32. Olorunshola KV, Achie LN, Akpomiemie ML. Ascorbic acid ameliorates toxic effects of chlopyrifos on testicular functions of albino rats. British Journal of Pharmacology and Toxicology. 2011;2(5): 262-269.
- 33. Mossa ATH, Abbassy MA. Adverse haematological and biochemical effects of certain formulated insecticides in male rats. Research Journal of Environmental Toxicology. 2012;6(4):160-168.
- Jaiswal C, Verma Y. Effect of alphatocopherol on haematological alteration induced by chlorpyrifos in albino rats. Haryana Veterinarian. 2012;51:108-110.
- 35. Ambali SF, Akanbi DO, Oladipo OO, Yaqub LS, Kawu MU. Subchronic chlorpyrifos- induced clinical, haematological and biochemical changes in swiss albino mice: Protective effect of

- vitamin E. Int. J. Biol. Med. Res. 2011; 2(2):497-503.
- 36. Hancock S, Ehrich M. Hinckley J. Pung T, Jortner BS. The effect of stress on the acute neurotoxicity of the organophosphate insecticide chlorpyrifos. (Special issue: Anticholinesterases). Toxicology and Applied Pharmacology. 2007;219(2/3):136-141.
- 37. USEPA U.S. Environmental protection agency. Health effects division chapter of the tolerance reassessment eligibility decision document; 2005.
- Bozkurt A, Yardan T, Ciftcjoglu E. Baydin A. Hakligor A. Bitigic M, Bilge S. Time course of serum S100B protein and neuron-specific enolase levels of a single dose of chlorpyrifos in rats. Basic and Clinical Pharmacology and Toxicology. 2010;107(5):893-898.
- 39. Johnson Frank O, Chambers Janice E, Nail Carole A. Givaruangsawat Sumalee, Carr L Russell. Developmental chlorpyrifos and methyl parathion exposure alters radial-arm maze performance in juvenile and adult rats. Toxicological Sciences. 2009;109(1):132-142.
- 40. Goel A, Dani V, Dhawan DK. Role of zinc in mitigating the toxic effects of chlorpyrifos on hematological alterations and electron microscopic observations in rat blood. Bio Metals. 2006;19(5):483-492.
- 41. Levine MS, Fox NL, Thompson B, Inhibition of esterase activity and an undercounting of circulating monocytes in a population of production workers. Journal of Occupational Medicine. 1986;28(3): 207–211.
- Corcoran GB, Fix L, Jones DP. Apoptosis: Molecular control point in toxicity, Toxicologyand Applied Pharmacology. 1994;128(2):169–181.
- 43. Rabideau CL. Pesticide mixtures induce immunotoxicity: Potentiation of apoptosis and oxidative stress, Thesis MS, Virginia Polytechnic Institute and State University, Blacksburg, Va, USA; 2001.
- Nakadai A, Li Q, Kawada T. Chlorpyrifos inducesapoptosis in human monocyte cell line U937. Toxicology. 2006;224(3):202– 209.
- 45. Thrasher JD, Madison R, Broughton A. Immunologic abnormalities in humans exposed to chlorpyrifos: Preliminary

- observations. Arch Environ Health. 1993; 48(2):89-93.
- 46. Galloway T, Handy R, Immunotoxicity of organophosphorous pesticides. Ecotoxicology. 2003;12(1-4):345-363.
- 47. Das BK, Mukherjee SC. Asian fisheries science Asian fisheries society. Manilla, Philippines. 2000;13:225-233.
- Olav AG, Ralf W, Axel MG. Biomarkers of liver fibrosis: Clinical translation of molecular pathogenesis based on liverdependent malfunction test. Clinica Chimica. Acta. 2007;381(2):107-113.
- 49. Rej R. Aminotransferase in disease, Clin. Lab. Med, 1989;9(4):667-87.
- Lawrence AK, Amadeo JP. (Ed). Clinical chemistry: Theory, analysis and correlation. The C.V. Mosby Company. 1989;367.
- 51. Mukut S, Ran S, Xiu LG. Brian LF. Ellen TM. Virginia JS, Eric PC, John EM. Early detection of radiation-induced glomerular injury by albumin permeability assay. Bio. One. 2001;155(3):474-480.
- 52. Luskova V, Svoboda M. Kolarova J. The effect of Diazinon on blood plasma

- biochemistry in carp (*Cyprinus carpio* L.). Acta Vet. Brno, 2002;71:117-123.
- 53. Jyotsna AP, Arun JP, Sanjay P. Biochemical effects of various pesticides on sprayers of Grape garden. Indian. J. Biochem. 2003;(18-21):16-22.
- Johnson KA. Toxicology and environmental research and consulting, midland, MI; Laboratory Project Study. 2005;1-35.
- USEPA US. Environmental protection agency. Health effects division chapter of the tolerance reassessment eligibility decision document. 2005.
- 56. El-Deeb AEA, Abd El-Aleem IM, Sherin SG. Harmful effect of some insecticides on vital parameters of albino rats. J. Egypt. Soc. Toxicol. 2007;36:53-60.
- 57. Revathi K, Kumar S. Histopathological alterations induced by sub acute dosage of chlorpyrifos 40% E.C. in Swiss albino mice J. Exper. Zool. India. 2000;3(1):45-47.
- 58. Lemus R. Abd El-Ghany A. Chlorpyrifos: An unwelcome pesticide in our homes. Rev. Environ. Health. 2000;15(4):421-33.

© 2017 Fayinminnu et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
http://sciencedomain.org/review-history/19572